

Effect of Extract of Fast Growing Species *Trifolium alexandrium* L. on Germination, Photosynthetic Pigments and Nutrient Uptake of *Prosopis cineraria* (L.) Druce

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ABSTRACT In a completely randomized design, the effect of the extract from *Trifolium alexandrium* (0, 0.2 and 0.4%) on the germination, some morphological characteristics, photosynthetic pigments, and nutrients uptake of *Prosopis cineraria* was evaluated. The highest germination rate and percentage (9.16 and 21%, respectively) were recorded in the 0.2% treatment. The maximum and minimum lengths of radicle (6.58 and 3.16 cm) and pedicel (6.56 and 14.23 cm), and dry weight were recorded in the control and 0.4% treatments, respectively. The highest level of chlorophyll a (16.80 mg g⁻¹ fresh weight) was found in the control. The highest and lowest chlorophyll b (9.65 and 7.96 mg g⁻¹ fresh weight) were measured in the 0.2% and control, respectively. The extract adversely affected the nutrient uptake by *P. cineraria*, the minimum and maximum of which were recorded at the 0.4% and control treatments, respectively. In general, although the extract of *T. alexandrium* increased the *P. cineraria* germination in the 0.2% treatment, the increased concentrations led to its reduced growth and nutrient uptake. Therefore, the simultaneous cultivation of these two species in the dry lands is not recommended.

Key words: Allelopathy, Plant growth, Plant extract, Photosynthetic pigments

1 INTRODUCTION

Allelopathy is one of the important issues in rehabilitation of rangelands, which has received less attention (Gholami *et al.*, 2011). Allelopathic effects, which can be seen in different plant species (Ricki Maryshany *et al.*, 2015), is a form of plant interference by means of interaction of plants with their released chemicals (Trezzi *et al.*, 2016), which play an important role in biodiversity and ecosystem functions (Iman and Zakaria, 2006). Various compounds in plants can affect various aspects

of plant communities (Narwal, 2004; Iman and Zakaria, 2006). Such compounds are classified as secondary plant materials or sub-materials of metabolic pathways of plants (Narwal, 2004), which are derived from the branches and leaves or secreted by the roots to the environment (Naseem *et al.*, 2009). Chemicals released by plants can act as natural herbicides or pesticides by controlling weeds (Narwal, 2004). Allelopathic studies on plants can, therefore, lead to discovery of new natural herbicides and growth inhibitors.

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There are more than 8000 plant species in Iran (Mozaffarian, 1996) with limited studies on their allelopathic properties. Leguminosae include important flowering plants with a wide distribution (Rechinger, 1984). *Prosopis cineraria* and *Trifolium alexandrinum* are two legume species found in Iran. *P.cineraria* is a native of the arid environments of several Western Asiatic countries (Bohra and Ghosh, 1980) and also found in extreme conditions with an annual rainfall as low as 15 cm and high alkaline and saline soils (Rechinger, 1984). The seed of *P. cineraria* has a hard germination (Manga and David, 1995; Sacheti and Al-Areimi, 2000). *T. alexandrinum* is a fast growing annual legume and one of the most important forage crops in the Mediterranean and the Middle-East regions (Hackney et al., 2007).

The most important strategy for reclamation and conservation of rangeland ecosystems is selecting suitable plant species. However, even with most adaptable species to be used for the rangeland, the improvement projects are likely to fail, if *allelopathy* properties are neglected (Bagheri and Mohammadi, 2011). Studies have shown that the germination and growth of *Chenopodium album* decreased under the allelopathic effect of *Atriplex nummularia* stem extracts (Bouchikh-Boucif et al., 2014). Sadaqa et al. (2010) also reported that the extract of *Amaranthus graesizans* had reduced the shoot and root dry weight of *Allium cepa* by 94% and 96%, respectively. The negative impacts of *Artemisia sieberi* on seed germination, stem and root of three important rangeland species has also been reported (Bagheri and Mohammadi, 2011). Similarly, Kazerooni Monfared et al. (2013) reported the negative effects of the aqueous extract of *T. alexandrium* shoot on seed germination of some weed species, and Hejazi et al. (2004) reported *T. alexandrium* extract decreased the growth of *Heliantus annuus*.

In the present study, the effect of *T. alexandrium* extract on the germination, morphological characteristics and nutrient uptake of *P. Cineraria* was studied, since it is one of the most important species in southern Iran and it has hard germination. Our hypothesis was that *T. alexandrium* extract acted as an enhancing growth on the germination, growth and nutrient uptake of *P. cineraria*.

2 MATERIALS AND METHODS

2.1 Plant extraction

The collected *T. alexandrium* samples were dried in the shade and ground to powder. Then, 190 g of the powder was put in a plastic bottle, then filled with 1 L ethanol and placed on a shaker for 24 hours. The resulting solution was filtered out and the extract was obtained using maceration method.

2.2 Preparation of pots

Soil was collected from the depths of 0–30 cm with a 5.5 cm diameter corer, then mixed, sieved to 4 mm, and its moisture content adjusted to 70% water-holding capacity (WHC). Soil characteristics were determined (Table 1); for texture, laser diffractometry was used (Wang et al., 2012), pH was measured in a 1:5 soil to distilled water slurry after 1 hour of agitation (Thomas, 1996), using a digital pH-meter (Model 691, Metrohm AG Herisau, Switzerland); for electrical conductivity (ECe), an EC-meter (DDS-307, Shanghai, China) was used (Rhoades, 1996); for the available phosphorus (AP) and potassium (AK), the methods of Bray and Kurtz (1954) and the flame photometry (Knudsen et al., 1982) were applied, respectively; the total N was analyzed calorimetrically using of Kjeldahl (Bremner, 1996).

Table 1 General characteristics of the soil used in the experiment

Texture	N (mg kg ⁻¹)	P (mg kg ⁻¹)	K (mg kg ⁻¹)	EC (dSm ⁻¹)	pH
Loamy sand	90	0.23	90	1.06	8.46

2.3 Germination medium

Plastic pots (diameter 10× diameter 15× height 45 cm) were filled with the sieved and dried soil. After disinfecting the seeds of *P. cineraria* with fungicide solution, 30 seeds were buried evenly throughout each pot, at least 3 cm from the edge. The pots were treated every two days with the extract of *T. alexandrium* (ml/L of distilled water) at concentrations of 0.2%, 0.4%, and 0% (only distilled water as control). The work was conducted in four replications under the greenhouse conditions (temperature 23±5°C, humidity 60%). The pots were irrigated with tap water as needed and the experiment was terminated 14 days after cultivation. The parameters measured included germination percentage and rate, radicle and pedicel length, seedling dry weight, photosynthetic pigments, and nutrient uptake by *P. cineraria*.

2.4 Calculation of plant properties

Germinated seeds were counted daily until the germination had been completed (Farajollahi *et al.*, 2012; Ebrahimi and Miri, 2016). On the last counting day (the 14th day), the radicle and pedicle lengths of 10 randomly selected seedlings from each pot were measured with a caliper, then washed with distilled water and placed in an oven (Dena-Iran) at 70 °C for 48 hr for drying, the weight of which was also measured. The germination rate and

germination percentage measured according to Eqs. 1 and 2.

$$GR = \sum Ni / Di \quad (1)$$

Where GR, Ni and Di were germination rate, number of germinated seeds in each day and: counted day, respectively (Maguire, 1962).

$$GP = (n/N) 100 \quad (2)$$

Where GP: germination percentage, n: total number of the germinated seeds during counting, N: total number of the germinated seeds in each pot (Behbodian *et al.*, 2005).

To measure chlorophyll a and b content, total chlorophyll, and carotenoids, 100 mg of fresh tissue was pulverized inside a porcelain mortar with 5 ml of 80% acetone, then centrifuged. The solution was transferred to centrifuge tubes, and the remnant in the mortar was washed twice with 5 ml of 80% acetone, the solution of which was added to the tubes. Then, the tubes were centrifuged for 10 min at 6000 rpm, the solution of which was transferred to a 250 mm flask, and its volume was adjusted to 25 ml with 80% acetone. Chlorophyll contents was read at wavelengths of 470, 663, 645 nm, using spectrophotometer (WPA-S2000) (Arnon, 1967). The contents of chlorophyll a, b and carotenoids were estimated according to the Eqs. 3, 4, 5. Total chlorophyll was calculated by sum of chlorophyll a and b in terms of milligrams per gram of sample weight (Arnon, 1967).

$$\text{Chlorophyll a} = 19.3 \times A_{663} - 0.86 \times A_{645} \quad (3)$$

$$\text{Chlorophyll b} = (19.3 \times A_{645} - 3.6 \times A_{663}) / V \quad (4)$$

$$\text{Carotenoides} = 100(A_{470} - 3.27 (\text{mgchl.a}) - 104 (\text{mgchl.b}) / 227 \quad (5)$$

Where V: volume of filtrated solution (upper solution of centrifuges), A: absorption of light

at wavelengths of 663, 645 and 470 nm, W: wet weight of sample (g).

2.4 Calculation of plant nutrient uptake

The wet oxidation method was used for the samples digestion to measure the absorbed elements. For this purpose, 0.3 g of the plant was transferred to digestion pipes, then 2.5 ml of a mixture of sulfuric acid, salicylic acid, selenium and hydrogen peroxide was added. The sample was shaken well and left for 2 hrs, then heated for 2 hr at 100 °C, and cooled. Then, one ml hydrogen peroxide was added and heated to 330 °C until the digestion was over when the extract turned colorless or pale yellow, which lasted for almost 2 hrs. After drying, 48.3 ml of distilled water were added to the pipes and, stirred. The next day, the stirring operation was repeated, and was put to itself to be deposited. Then, the elements of N, P, K, Zn, Mn were measured (Einhelling and Leather, 1988). The amount of Mn and Zn were measured using atomic absorption spectrophotometry (GBC Avanta, Australia). Nitrogen was measured using Kjeldahl method (Gerhardt 9801/Ac), phosphorus through the colorimetric method by a spectrophotometer (JENWAY 640), and K was measured using a flame photometer (Ryan *et al.*, 2001).

2.5 Statistical analysis

The data were statistically analyzed using the SAS 8.1. The statistical processing was mainly conducted by analysis of variance (ANOVA), and the normality of data was tested using

Kolmogorov–Smirnov. When needed, log transformation of the data was conducted to achieve normality. Equality of variance among treatments was tested using Levene's test for homogeneity of variance. Duncan test post hoc analysis was performed to define which specific mean pairs were significantly different.

3 RESULTS

3.1 Effect on germination and morphological properties

The extract of *T. alexandrium* significantly affected the germination rate and percentage of *P. cineraria*, the highest of which were recorded for the 0.2% treatment, while the lowest were found in the control treatment (Table 2).

The extract of *T. alexandrium* also significantly affected the radicle and pedicle lengths of *P. cineraria*. Increased concentration of the extract was associated with the decreased lengths of the radicle and pedicle (Table 2). The highest radicle and pedicle lengths were recorded in the control treatment, while the minimum lengths were found in the 0.4% extract treatment. The dry weight of *P. cineraria* was also significantly reduced with the increased concentration of the extract. The maximum dry weight was measured in the control treatment, while the lowest dry weight was found in the 0.4% treatment (Table 2).

Table 2 Germination and morphological properties of *P. cineraria* treated with *T. alexandrium* extract

Extract (%)	Germination rate	Germination percentage	Radicle length (cm)	Pedicle length (cm)	Total dry weight (g pot ⁻¹)
Control	7.00 ± 0.51c	33.3 ± 2.50c	6.58 ± 0.40a	14.23 ± 1.20a	0.86 ± 0.01a
0.2	21.00 ± 2.22a	91.6 ± 4.3a	5.23 ± 0.40b	12.27 ± 1.75a	0.76 ± 0.01b
0.4	12.00 ± 1.13b	58.3 ± 2.30b	3.16 ± 0.30c	6.56 ± 1.09b	0.54 ± 0.02c

*Values within a column followed by the different letters are significantly different (P < 0.05, means ± SE).

3.2 Effect on photosynthetic pigment

The photosynthetic pigments of *P. Cineraria* were affected under *T. alexandrium* extract treatment, however inconspicuously. The chlorophyll was inversely affected with the increased extract concentrations, so that the lowest level of chlorophyll a was recorded in the 0.4% extract treatment (Table 3), which was significantly different from the control

treatment. The highest level in chlorophyll b was found in the 0.2% treatment, followed by the 0.4%. Overall, the highest level in the total chlorophyll was recorded in the 0.2% treatment. The highest and lowest levels of carotenoids were related to 0.4% and the control treatments, respectively (Table 3).

Table 3 Photosynthetic pigments of *P. cineraria* treated with *T. alexandrium* extract

Extract (%)	Chlorophyll a (mg g ⁻¹ fresh weight)	Chlorophyll b (mg g ⁻¹ fresh weight)	Total Chlorophyll (mg g ⁻¹ fresh weight)	Carotenoid (mg g ⁻¹ fresh weight)
Control	16.80±2.40a	7.96±1.10c	24.577±1.40b	1.18±0.09b
0.2	15.56±2.31a	16.79±1.43a	32.34±2.40a	1.27±0.09b
0.4	12.48 ±2.01b	9.65 ±1.40b	22.13±2.30b	3.51±0.50a

*Values within a column followed by the different letters are significantly different (P<0.05, means±SE).

3.3 Effect on nutrient uptake

The extract of *T. alexandrium* affected the nutrient (N, P, K, Zn and Mn) uptake by *P. cineraria*, although inconspicuously (Table 4). Differences in N, Mn and Zn contents were non-significant between the control and 0.4% treatments. Potassium content showed non-

significant difference between the 0.2 and 0.4% treatments. In general, the maximum and minimum nutrient uptakes were recorded in the control and 0.4% extract treatments, respectively.

Table 4 Nutrient uptake by *P. cineraria* treated with *T. Alexandrium* extract

Extract (%)	N (mg g ⁻¹)	P (mg g ⁻¹)	K (mg g ⁻¹)	Zn (mg g ⁻¹)	Mn (mg g ⁻¹)
Control	2.75±0.03b	0.43±0.03a	0.43±0.01a	0.13±0.01b	0.17±0.01b
0.2	3.53±0.03a	0.30±0.03b	0.23±0.00b	0.16±0.01a	0.18±0.01a
0.4	2.86 ±0.03b	0.23 ±0.00c	0.21±0.00b	0.12±0.01b	0.15±0.01b

*Values within a column followed by the different letters are significantly different (p<0.05, means±SE).

4 DISCUSSIONS

Decomposition of plant in the soil have been shown to have allelopathic effects by releasing compounds such as phenolic acids that can have negative effects on the germination or growth performance in some plants (Purvis *et al.*, 1985; Hoffman *et al.*, 1996; Naseem *et al.*, 2009). These effects are selective and depend on the

concentration and type of residue and, may lead to stimulating the growth inhibitory effects in plants. Low concentration of *T. alexandrium* extract had a positive effect on the germination of *P. cineraria*. Because the allelopathic phenomenon much depends on the concentration of the allelochemicals, and any change in the amount of these materials leads to

different inhibition and stimulation effect (Chon and Kim, 2002; Koloren, 2007). It is interesting to note that allelopathic substances in low concentrations may have positive or negative effects on the target species, but high concentrations are always associated with ambiguous changes (Ismail and Chong, 2002). The reason for reduced germination at 0.4% extract can be related to the enzyme activities such as amylase that plays an important role in seed germination (Soltanipor *et al.*, 2007; Bagheri and Mohammadi, 2011). The results indicate that allelopathic compounds may but not sure reduce plant germination with effect on hormones, such as gibberellin, which is important in plant germination, as well as the effect on the activity of special enzymes, such as amylase and proteinase, which are essential in the process of germination. In addition, reduction in the germination stage may be attributed to change in the activity of enzymes that affect the transfer of storage compositions during germination. Inhibiting substances secreted from different plant organs, may result in the accumulation of phenolic compounds and reduced germination percentage (Ghorbanli *et al.*, 2008).

Accumulation of phenolic compounds is considered as a defense response against biotic and abiotic stresses that reduces the plant growth and germination (Hirt and Shinozaki, 2004). For example, germination percentage of some weeds species (Kazerooni Monfared *et al.*, 2013) and sunflower (Hejazi *et al.*, 2004) were significantly affected when treated with the extract of *T. alexandrium*. In addition, Ghorbanli *et al.* (2008) reported that germination percentage of *Avena ludoviciana* was significantly affected by the extract of *Artemisia* extract.

SafwanIshak *et al.* (2016) found decreased germination, root growth and fresh weight of some plants with increased concentrations of extract from *L. leucocephala*.

The results of the present study showed that increase in the concentration of *T. alexandrium* extract was associated with decrease in lengths of the radicle and pedicle as well as reduced dry weight of *P. cineraria*. The reduced radicle and pedicle lengths may be due to hormonal balance and, reduced shoot growth. Some mechanisms of allelopathic activities are similar to plant hormones. *Decreased* seedlings' length of plant which are exposed to allelopathic compounds may be due to the negative effect of the extract on cell division or cell elongation, which in addition to longitudinal growth of the plant, inhibiting substances in extract can have a negative impact on weight plant (Qasem, 2001). Root is the first part that absorbs allelopathic materials directly from the environment, and so compared to other traits may be more affected (Kazerooni Monfared *et al.*, 2013). This is due to the fact that the impact on root growth reduces water absorption and, thereby, reduces the seedlings' length (Chon *et al.*, 2002). Reductions in the root growth under allelopathic stress in some plants have reportedly been higher than the shoot growth (El-Khatib *et al.*, 2016). Increased antagonistic effects as the result of increase in the extract concentration have also been reported (Saraei *et al.*, 2011; Samedani and Baghestani, 2005; Gong *et al.*, 2016).

One reason for the reduced growth rate during allelopathic stress has been attributed to changes in the mitochondrial respiration rate that, in turn, decreases the ATP production, which can cause changes in other cellular processes, such as ion adsorption and growth. Decreased plant growth in the presence of allelopathic compounds is associated with the reduced mitosis of root and shoot's meristem cells and therefore the length of root and shoot will be reduced (Bertin *et al.*, 2003).

The effects of allelopathic materials on the photosynthetic pigments have already been demonstrated (Hejazi, 2001; Ricki Maryshany

et al., 2015). In the present study, *T. alexandrium* extract showed negative effect on the contents of photosynthetic pigments in *P. cineraria*. The reason for the reduced chlorophyll contents at high concentrations may be decomposition of chlorophyll and carotenoid or their reduced synthesis (Chaniago *et al.*, 2006; Ricki Maryshany *et al.*, 2015). The decreased chlorophyll contents as the result of allelochemical may be a secondary effect (Babu and Kandasamy, 1997).

Nutrient uptake is an important factor for plant growth and development, the rate of which can be affected by the accumulation of allelopathic factors (Mallik, 2005; Mohammaddoust Chamanabad *et al.*, 2014). Both increases and decreases in nutrient uptake have been reported for plants that are subjected to the allelopathic conditions. Unstable situation of minerals in receiver plants is created by leaching of plant debris, root exudates and allelopathic debris, the effects of which may directly be related to plants competition, and indirectly through microorganisms that stabilize the nutrients (Alam *et al.*, 2001). Special allelochemical (flavonoids and phenolic acids) prevent the minerals uptake through disrupting the normal actions of membrane in the root cells. Allelochemical can reduce cellular ATP content through inhibition of electron transport and oxidative phosphorylation, which are two mitochondrial membrane actions, as well as change the membrane permeability to inorganic ions uptake (Bhowmik and Doll, 1984). Yu and Matsui (1997) found that the root exudates of cucumbers and their analogues prevented the uptake of $H_2PO_4^-$ by the seedlings. Mohammaddoust Chamanabad *et al.* (2014) reported that fresh and dry extract of *Sinapisarvensis* and *Cirsiumarvense* increased K concentrate in the tested plant.

5 CONCLUSION

In the present study, extract of *T. alexandrium* influenced all traits of *P. cineraria*. However, the lower concentration had positive effect on the germination percentage. Given the positive impact of *T. alexandrium* on increasing germination of *P. cineraria*, the former plant can be used to increase the germination of *P. cineraria* that has germination problem. But given the negative impact of *T. alexandrium* on the growth and nutrient uptake in of *P. cineraria*, cultivation of two plants is not recommended in arid rangelands.

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7 REFERENCES

- Alam, S.M., Ala, S.A., Azmi, A.R., Khan, M.A. and Ansari, R. Allelopathy and its role in agriculture. J. Bio.Scie., 2001; 1: 308-315.
- Arnon, D.I. Copperenzymes in isolated chloroplasts. Polyphenol oxidase in *Beta vulgaris* Plant Physiol., 1949; 24: 1-5.
- Babu, R.C. and Kandasamy, O.S. Allelopathic effect of *Eucalyptus globulus* Labill. On *Cyperusrotundus* L. and *Cynodondactylon* L. Pers. J. Agron. Crop. Sci., 1997; 179: 123-126.
- Bagheri, R. and Mohammadi, S. Allelopathic effects of *Artemisia sieberi* Besser on three important species (*Agropyrondesertorum*, *Agropyronelongatum* and *Atriplexcanescens*) in range improvement. Iran J. Range and Desert Res., 2011; 17: 538-549. (In Persian).
- Behbodan, B., Lahouti, M. and Nezami, A. Effects evaluation of salt stress on germination of chickpea varieties. Agron. J., 2005; 28: 127--137. (In Persian).

- Bertin, C., Yang, X. and Weston, L.A. The role of root exudates and allelochemicals in the rhizosphere. *Plant Soil*, 2003; 256:67-83.
- Bhowmik, P.C. and Doll, J.D. Allelopathy effect of annual weed residues on growth and nutrient uptake of corn and soybeans. *Agron. J.*, 1984; 76:383-388.
- Bohra, M.C. and Ghosh, P.K. The nutritive value and digestibility of *Prosopis cineraria* in Khejri in the Indian Desert. *Cazri-Icar.*, 1980; 45-47.
- Bouchikh-Boucif, Y, Labani, A., Bebabdeli, K.H. and Bouhelouane, S. Allelopathic effects of shoot and root extracts from three alien and native *Chenopodiaceae* species on lettuce seed germination. *Ecologia Balakanica.*, 2014; 2: 51-55.
- Bray, R.H. and Kurtz, L.T. Determination of total, organic and available forms of phosphorus in soils. *Eur. J. Soil Sci.*, 1954; 39-45.
- Bremner, J.M. Nitrogen total, *Methods of Soil Analysis*, Bartels, J.M. (Ed.), Soil Sci. Soci. Am., Madison, Wisconsin. 1996; 1085-1122.
- Chaniago, I., Taji, A. and Jessop, R. Weed interference in soybean (*Glycine max*). *Proceedings of the 13th Australian agronomy conference*. Perth, Australia., 2006; 542-544.
- Chon, S.U. and Kim, J.D. Biological activity and quantification of suspected allelochemicals from Alfalfa plant parts. *J. Agron. Crop Sci.*, 2002; 188: 281-285.
- Chon, S.U, Choi, S.K., Jang, H.G., Pyo, B.S. and Kim, S.M. Effects of alfalfa leaf extracts and phenolic allelochemicals on early seedling growth and root morphology of alfalfa and barnyard grass. *Crop Prot.*, 2002; 21:1077-1082.
- Ebrahimi, M. and Miri, E. Effect of humic acid on seed germination and seedling growth of *Borago officinalis* and *Cichorium intybus*. *ECOPERSIA*, 2016; 4: 1239-1249. (In Persian)
- Einhelling, F.A. and Leather, G.R.. Potential for exploiting allelopathy to enhance crop production. *J. of Chemi. Ecol.*, 1988; 4: 1829-1844.
- El-Khatib, A.A., Barakat, N.A. and Nazeir, H. Growth and physiological response of some cultivated species under allelopathic stress of *Calotropis procera* (Aiton) W.T. *App. Sci. Rep.*, 2016; 14: 237-246.
- Farajollahi, A., Tavili, A., Gholinejad, B., Darini, J. and Pouzesh, H. Investigation and compare the allelopathic effects for different tissues of *Peganum harmala* in different amounts on the *Bromus tectorum* germination and growth characteristics. *ECOPERSIA*, 2012; 1: 217-226. (In Persian)
- Gholami, P., Ghorbani, J. and Ghaderi, Sh. Allelopathic effects of *Artemisia aucheri* on seed germination and *Dactylis glomerata* properties of *Festuca arundinacea* Schreb. *Plant Eco-physiology.*, 2011; 9: 41-52. (In Persian)
- Ghorbanli, M.L., Bakhshi Khanegi, G.R. and Shojaie, A.A. Survey of allelopathic potential of *Artemisia siberi beaser* on *Avena lodoviciana* seedling and *Amaranthus retroflexus*. *Pajouhesh and Sazandegi*, 2008; 129-134. (In Persian).
- Gong, Z., Fu, X., Zhou, X., Wu, F. and Liu, S. Effects of plant residues on cucumber (*Cucumis sativus* L.) growth, soil enzyme activities and microbial communities. *Allelopat. J.*, 2016; 38: 147-158.
- Hackney, B., Dear, B. and Crocker, G. Berseem clover. New South Wales Department of Primary Industries, Primefacts. 2007. 388p.
- Hejazi, A, Ghaffari, S.M. and Hosseini Mazinani, S.M. effect allelopathic possible Roots of wheat, cotton and

- sunflower on different stages of development Sunflower seed yield. Res. Devel., 2004; 51: 88-93.
- Hejazi, A. Allelopathy (Autotoxicity and Hetrotoxicity. University of Tehran press, 2001; 181p. (In Persian).
- Hirt, H. and Shinozaki, K. Plant responses to abiotic stress topics in current genetics. Springer Verlag, Berlin, Germany, 2004; 297p.
- Hoffman, M.L., Weston, L.A., Shyder, J.C. and Regnier, E.E. Separating the effects of sorghum (*Sorghum bicolor*) and rye (*Secale cereal*) root and shoot residues on weed development. Weed Sci., 1996; 44: 402-407.
- Iman, A. and Zakaria, W. Allelopathic effect of sweet corn and vegetable soybean extracts at germination and seedling growth of corn and soybean varieties. Agron. J., 2006; 5: 62-68.
- Ismail, B.S., and Chong, T.V. Effect of aqueous extract and decomposition of *Mikania micrantha* on selected agronomic crops. Weed Biol. Manag., 2002; 2: 31-38.
- Kazerooni Monfared, S., Tokasi, M. and Banayan Awal, M. Study of allelopathic effects of berseem clover (*Trifolium alexandrium*) shoot aqueous extract on germination and initial seedling growth of some weed species. J. Plant Protection., 2013; 27: 509-512. (In Persian).
- Knudsen, D., Peterson, G.A. and Pratt, P. Lithium, Sodium and Potassium, Methods of Soil Analysis, Page, A.L. (Ed.), Am. Soci. Agron., Madison, Wisconsin. 1982; 225-246.
- Koloren, Q. Allelopathic effects of *Medicago sativa* L. and *Viciacracca* L. leaf and root extracts on weeds. Pak J Biol Sci., 2007; 10: 1639-1642.
- Maguire, J. Speed of germination-aid in selection and evaluation for seedling emergence and vigour. Crop Sci., 1962; 2: 176-177.
- Mallik, A.U. Allelopathy: advances, challenges and opportunities. Proceedings of the 4th world congress on allelopathy. Charles Sturt University, Wagga, NSW, Australia. International Allelopathy Society. 2005: 3-11.
- Manga, V.K. and David, N.S. Influence of seed traits on germination in *Prosopis cineraria* (L.) MacBride. Arid Environ., 1995: 371-375.
- Mohammaddoust Chamanabad, H.R., Sayaah, M., Asghari, A. and Pourmorad Kaleibar, B. The allelopathic effects of fresh and dry residual extract of Wild mustard (*Sinapisarvensis*) and Canada Thistle (*Cirsiumarvense*) on germination and nutrient uptake of canola (*Brassica napus*). Pajouhesh and Sazandegi., 2014; 104: 41-47. (In Persian).
- Mozaffarian, V.A. Dictionary of Iraninan Plant Names. Farhang Mo'aser, 1996; 566p. (In Persian).
- Narwal, S. Allelopathy in Crop Production. Jodhpur, Scientific Publishers, 2004; 303p.
- Naseem M., Aslam, M., Asnar, M. and Azhar, M. Allelopathic effects of sunflower water extract on weed control and wheat productivity. Pak. J. Weed Sci. Res., 2009; 15: 107-116.
- Nelson, E.B. Effects of Allelochemicals on Mineral Uptake and Associated Physiological Processes. ACS Symposium Series, 1985; 268p.
- Purvis, C.E., Jessop, R.S. and Lovea, J.V. Selective regulation of germination and growth of annual weeds by crop residues. Weed Res., 1985; 25: 415-421.

- Qasem, J.R. Allelopathic potential of white top and Syrian sage on vegetable crops. *Agron. J.*, 2001; 93: 64-71.
- Rechinger, K.H. 1984. *Flora Iranica. Papilionaceae*, Akademische Druck U Verlagsanstalt, Graz, Austria.
- Rhoades, J.D. Salinity: Electrical conductivity and total dissolved solids, *Methods of soil analysis*, Page, A.L. (Ed.). Soil Science Society of America, Madison: Wisconsin. 1996; 417-435.
- Ricki Maryshany, A. Ebrahimi. And Shirmohammadi, E. Effects of *Trifolium alexandrinum*, *Artemisia sieberi* and fertilizer on agree extract on germination and morphological properties of *Peganumharmala* and *Prosopis cineraria*. Range Management M.Sc. Thesis. University of Zabol, 2015; 86p. (In Persian)
- Ryan, J., Estefan, G. and Rashid, A. *Soil and Plant Analysis Laboratory Manual*, International Centre for Agricultural Research in the Dry Areas (ICARDA). Aleppo and National Agricultural Research Centre (NARC), Islamabad, Pakistan. 2001; 172p.
- Sacheti, U and Al-Areimi. The influence of high storage and germination temperature on the germination of *Prosopis cineraria* seeds from northern Oman. *J. Tro. For. Sci.*, 2000; 191-193.
- Sadaqa, E.A., Bawazir, A.A. and Qasem, J.R. Allelopathic activity of some common weeds species in onion fields. *Allelopat. J.*, 2010; 26: 175-184.
- SafwanIshak, M., Ismail, B.S. and Yusoff, N. Allelopathic potential of *Leucaenaleucocephala* (Lam.) de Wit on the germination and seedling growth of *Ageratum conyzoides* L., *Tridax procumbens* L. and *Emilia sonchifolia* (L.) DC., *Allelopat. J.*, 2016; 37: 109-122.
- Samedani, B. and Baghestani, M.A. Comparison of allelopathic activity of different *Artemisia* species on seed germination rate and seedling growth of *Avenaludoviciana*. Pajouhesh and Sazandegi., 2005; 68: 69-74. (In Persian).
- Saraei, R., Lahouti, M. and Ganjeali, A. Evalution of allelopathy effects of eucalyptus (*Eucalyptus globulus* Labill.) on germination, morphological and biochemical of barley (*Hordeumvulgare*) and flix weed (*Descurainia Sophia*). *Agroecology*.2011; 4: 215-222.(In Persian).
- Soltanipor, M., Hajebi, A., Dastjerdi, A. and Ebrahimi, S. Allelopathic effects of aqueous extract of *Zhumeri amajdaeon* seed germination of seven species of vegetables. *Iran J. Med. Aromat .Plants.*, 2007; 23: 51-58.
- Thomas, G.W. Soil pH and soil acidity. *Methods of soil analysis*, Sparks D.L. (Ed.), Am. Soci. Agron. Soil Sci. Madison: Wisconsin., 1996; 475-490.
- Trezzi, M.M., Vidal, R.A., Balbinot Junior, A.A., HertwigBittencourt, H.V. and Silva Souza Filho, A.P. Allelopathy: driving mechanisms governing its activity in agriculture. *J. Plant Interact.*, 2016; 1:53-60.
- Wang, A., Luo, C., Yang, R., Chen, Y., Shen, Z. and Li, X. Metal leaching along soil profiles after the EDDS application –A field study. *J. Environ. Pollut.*, 2012; 164: 204-210.
- Yu, J.Q. and Matsui, Y. Effect of root exudates of cucumber (*Cucumis sativus*) and allelochemicals on ion uptake by cucumber seedlings. *J. Chem. Ecol.*, 1997; 23: 817-827.

تاثیر عصاره گونه سریع‌الرشد *Trifolium alexandrium* L. در جوانه‌زنی، رنگیزه‌های فتوسنتزی

و جذب عناصر

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چکیده: در یک طرح کاملاً تصادفی با چهار تکرار، تاثیر عصاره *Trifolium alexandrium* در مقادیر صفر (کنترل)، ۰/۲، ۰/۴ درصد بر جوانه‌زنی، برخی خصوصیات موفولوژیک، رنگیزه‌های فتوسنتزی و جذب عناصر غذایی توسط *Prosopis cineraria* بررسی شد. بیش‌ترین درصد و سرعت جوانه‌زنی (به ترتیب ۹/۱۶ و ۲۱ درصد) در غلظت ۰/۲ درصد محاسبه شد. بیش‌ترین و کم‌ترین طول ریشه‌چه (۶/۵۸ و ۳/۱۶ سانتی‌متر)، ساقه‌چه (۱۴/۲۳ و ۶/۵۶ سانتی‌متر) و وزن خشک گیاه به ترتیب در تیمار شاهد و غلظت ۰/۴ درصد اندازه‌گیری شد. بیش‌ترین مقدار کلروفیل a (۱۶/۸۰ میلی‌گرم بر گرم وزن تر) مربوط به شاهد بود. بیش‌ترین و کم‌ترین کلروفیل b (۹/۶۵ و ۷/۹۶ میلی‌گرم بر گرم وزن تر) در تیمار ۰/۲ درصد و تیمار شاهد اندازه‌گیری شد. عصاره در مقایسه با تیمار شاهد باعث کاهش جذب نیتروژن، پتاسیم، فسفر، روی و منگنز توسط بافت‌های *P. cineraria* گردید. کم‌ترین و بیش‌ترین مقدار جذب عناصر غذایی به ترتیب در غلظت ۰/۴ عصاره و تیمار شاهد به دست آمد. به طور کلی هرچند عصاره *T. Alexandrium* در غلظت ۰/۲ درصد باعث افزایش جوانه‌زنی *P. cineraria* شد، اما افزایش غلظت، منجر به کاهش رشد و جذب عناصر غذایی گردید و کشت این دو گونه گیاهی باهم در مراتع خشک توصیه نمی‌شود.

کلمات کلیدی: دگرآسیبی، رشد گیاهان، عصاره گیاهی، رنگدانه‌های فتوسنتزی